

# Estadística para Bioinformática

...experiencia de un usuario no-estadista

## Corrección por múltiples comparaciones: valores p ajustados.

- 1. RNA-seq – p-value
- 2. Differentially expressed genes - FDR
- 3. Epigenetics – Fischer test
- 4. GO Terms – Hypergeometric test

# Identifying Differentially Expressed Genes

- Individual Hypotheses Testing

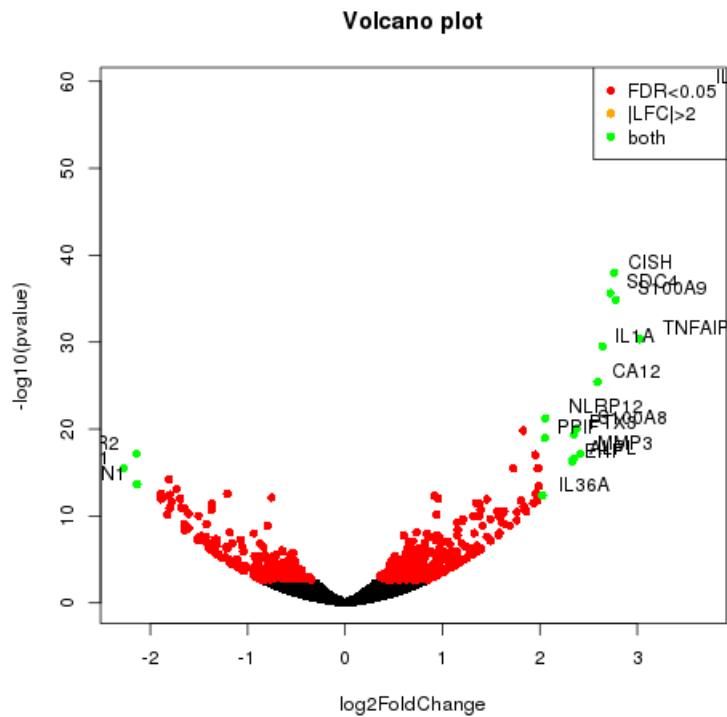
For each gene use a significance test of 0.05 level

$H_0$ : Gene is similarly expressed

$H_1$ : Gene is differently expressed

A t-statistic is calculated for comparing gene expression mean between the control and treatment groups.

## RNA-set output



P-value?

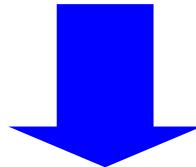
Thousands of genes in an experiment



Thousands of hypotheses are tested



Probability of type I error (false discovery)  
committed increases sharply



Multiplicity problem!

Are you sure all the tests are independent?

Multiple Testing Matters! Is actually essential

Genomics = Lots of Data = Lots of Hypothesis Tests

*A typical RNA-seq experiment might result in performing 6,000/25,000 separate hypothesis tests. If we use a standard p-value cut-off of 0.05, we'd expect ? genes to be deemed "significant" by chance.*

$$P(\text{Making an error}) = \alpha$$

$$P(\text{Not making an error}) = 1 - \alpha$$

$$P(\text{Not making an error in } m \text{ tests}) = (1 - \alpha)^m$$

$$P(\text{Making at least 1 error in } m \text{ tests}) = 1 - (1 - \alpha)^m$$

Experiments in finding people with paranormal powers:  
Joseph Rhine (1950)

**1000 people guessed the sequence of 10 cards: red or black?**



**12 persons guessed 9 of 10 cards, two of them all 10 cards**

**All these “physics” in further experiments didn’t confirm their paranormal abilities**

# What did really happen?

$$\text{Probability to guess all 10 cards} = \left(\frac{1}{2}\right)^{10} \approx 0.00098$$

$$\text{Probability to guess 9 cards} = 10 \left(\frac{1}{2}\right)^{10} \approx 0.0098$$

$$\text{Probability to guess 9 or all 10 cards} = 11 \left(\frac{1}{2}\right)^{10} \approx 0.0107$$

$$\begin{aligned} &\text{Chances to find a “psychic”} \\ &\text{among 100 persons} = 1 - (1 - 0.0107)^{100} \approx 0.66 \end{aligned}$$

$$\begin{aligned} &\text{Chances to find a “psychic”} \\ &\text{among 1000 persons} = 1 - (1 - 0.0107)^{1000} \approx 0.9998 \end{aligned}$$

# How to avoid false discovery

During  $m$  independent statistic test with  $\alpha$  significance level, the probability of at least one false discovery should be

$$1 - (1 - \alpha)^m < 0.05$$

$$\alpha = 1 - (1 - 0.05)^{1/m} \approx \frac{0.05}{m}$$

Bonferroni correction: during  $m$  independent statistic tests only those results are significant, for which

$$p < \frac{0.05}{m}$$

## What Does Correcting for Multiple Testing Mean?

- When people say “adjusting p-values for the number of hypothesis tests performed” what they mean is **controlling the Type I error rate**
- Very active area of statistics - many different methods have been described
- Although these varied approaches have the same goal, they go about it in fundamentally different ways

## The False Discovery Rate (FDR) criterion

Benjamini and Hochberg (95) :

$R$  = # rejected hypotheses = # discoveries

$V$  of these may be in error = # false discoveries

The error (type I) in the entire study is measured by

$$Q = \begin{cases} \frac{V}{R} & R > 0 \\ 0 & R = 0 \end{cases}$$

i.e. the proportion of false discoveries  
among the discoveries (0 if none found)

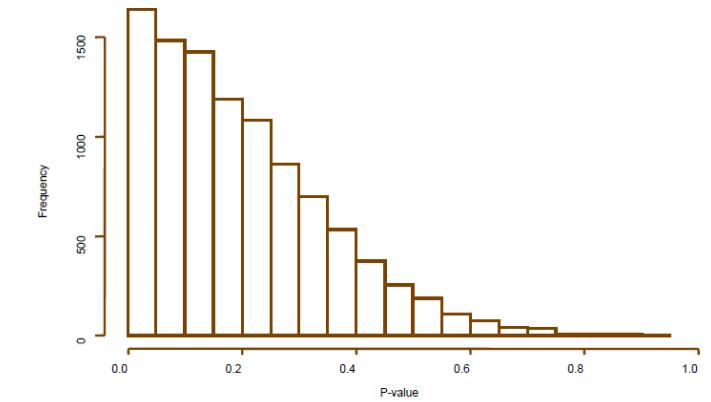
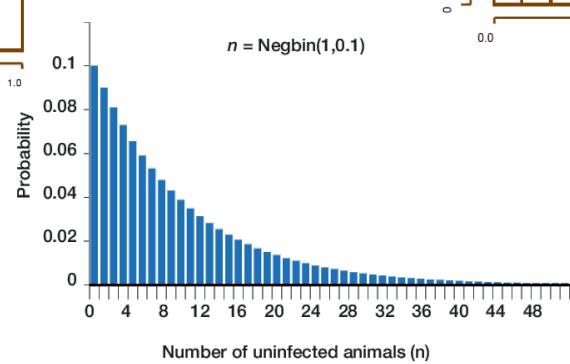
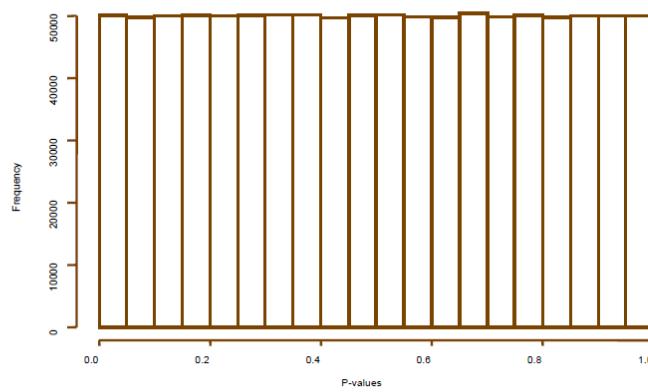
$$FDR = E(Q)$$

Does it make sense?

## What's a q-value?

- q-value is defined as the minimum FDR that can be attained when calling that “feature” significant (i.e., expected proportion of false positives incurred when calling that feature significant)
- The estimated q-value is a function of the p-value for that test and the distribution of the entire set of p-values from the family of tests being considered (Storey and Tibshirani 2003)
- Thus, in an array study testing for differential expression, if gene X has a q-value of 0.013 it means that 1.3% of genes that show p-values at least as small as gene X are false positives

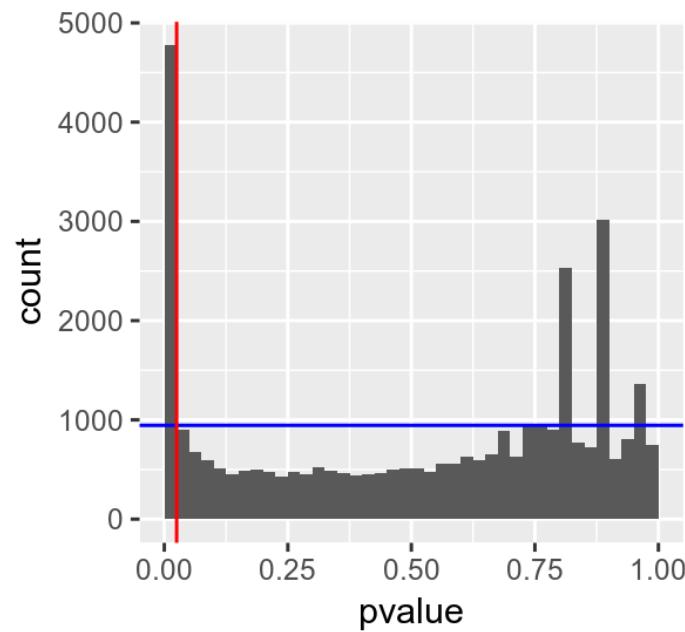
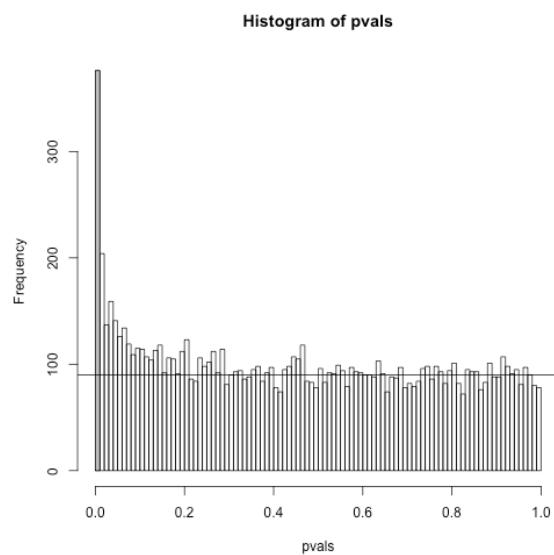
- Under the null hypothesis p-values are expected to be uniformly distributed between 0 and 1
- Under the alternative hypothesis p-values are skewed towards 0



Negative binomial distribution

# FDR

False discovery rate (FDR) is the expected proportion of Type I errors among the rejected hypotheses



# Example

```
#Install qvalue package
if (!requireNamespace("BiocManager", quietly = TRUE))
  install.packages("BiocManager")

BiocManager::install("qvalue")

#call package
library(qvalue)

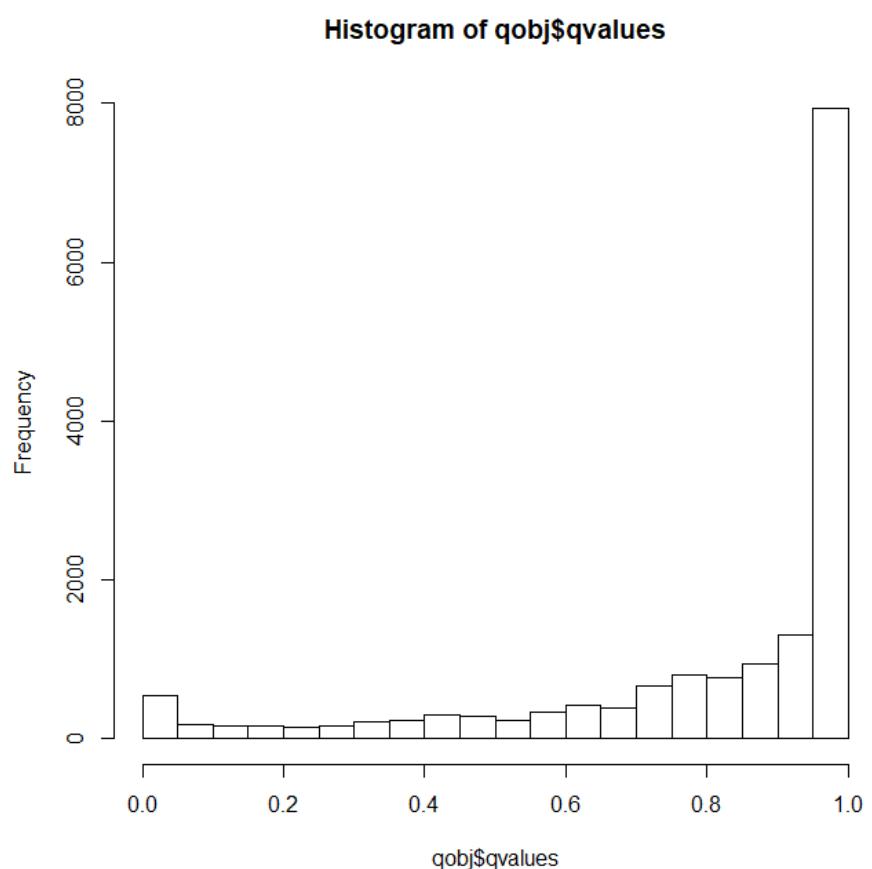
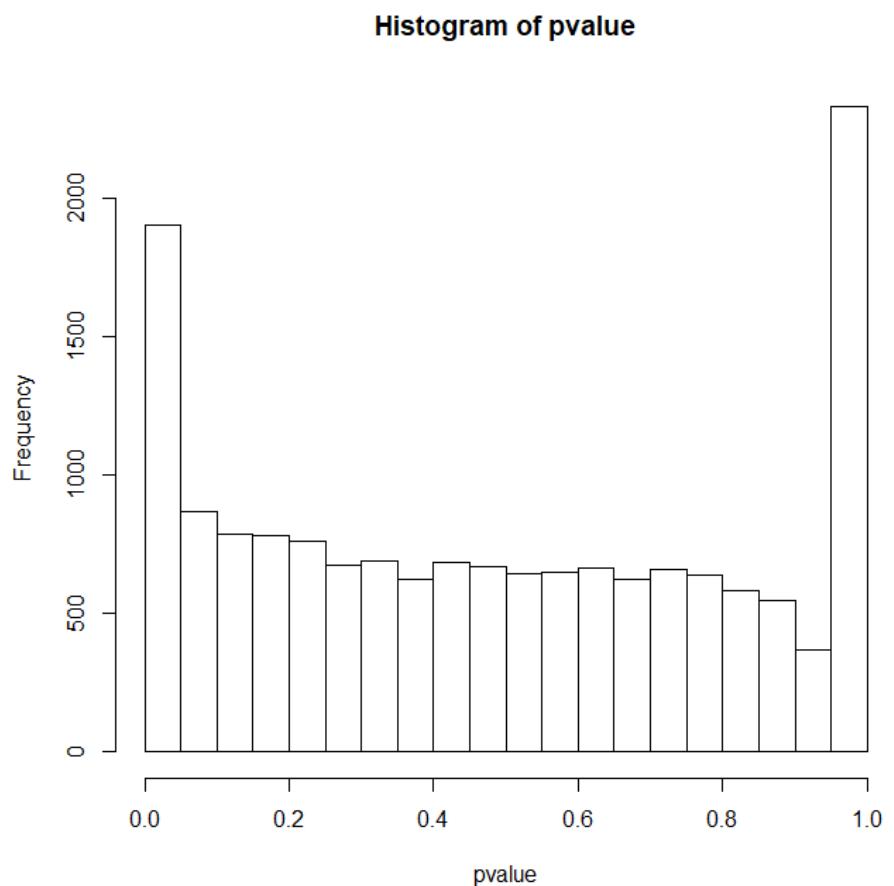
#add your file
pvalue <- scan("C:\\\\Users\\\\Usach\\\\Desktop\\\\Pvalues.csv")
head(pvalue)

#visualise the data distribution
hist(pvalue)

#run q_value

qobj <- qvalue(pvalue, fdr.level=0.05)
summary(qobj)
hist(qobj$qvalues)
plot(qobj)

#save pdf
pdf("plot_q_values.pdf")
plot(qobj)
dev.off()
```



# Criticism of FDR approach

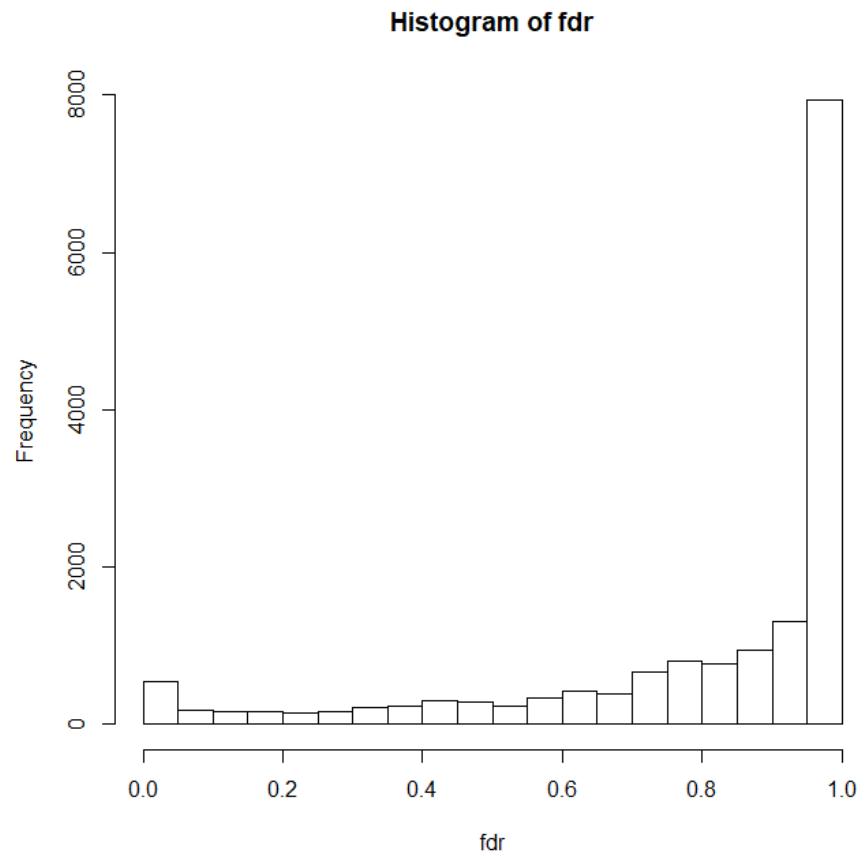
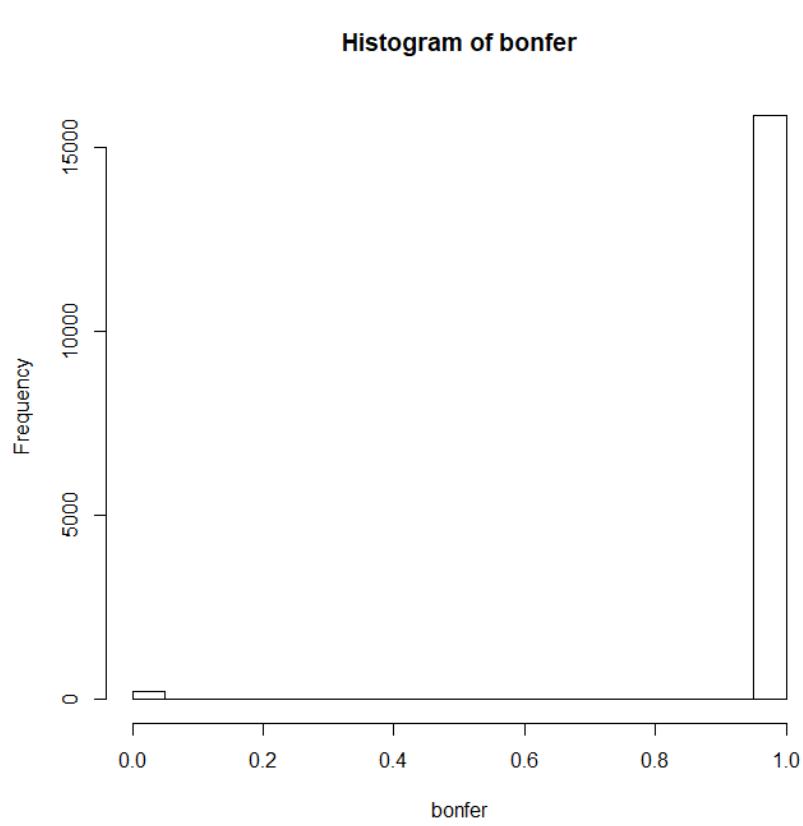
- It is "possible to cheat":
  - You can choose what p-values to use to alter the distribution and the outcome

## Bonferroni vs FDR

*Bonferroni: Very simple method for ensuring that the overall Type I error rate of  $\alpha$  is maintained when performing  $m$  independent hypothesis tests*

```
bonfer <- p.adjust(pvalue, method = 'bonferroni', n = length(pvalue))
head(bonfer)
summary(bonfer)
hist(bonfer)

fdr <- p.adjust(pvalue, method = 'fdr', n = length(pvalue))
head(fdr)
summary(fdr)
hist(fdr)
matplot(pvalue, fdr)
matplot(pvalue,bonfer)
```



## Epigenetics – Fisher test/chi-square

### What is Fisher's Exact Test of Independence?

Fisher's Exact Test of Independence is a statistical test used when you have two nominal variables and want to find out if **proportions** for one nominal variable are different among values of the other nominal variable. For experiments with small numbers of participants (under around 1,000), Fisher's is more accurate than the [chi-square](#) test or G-test.

A **chi-square test for independence** compares two variables in a contingency table to see if they are related. In a more general sense, it tests to see whether distributions of [categorical variables](#) differ from each another.

- A **very small chi square test statistic** means that your observed data fits your expected data extremely well. In other words, there is a relationship.
- A **very large chi square test statistic** means that the data does not fit very well. In other words, there isn't a relationship.

## When to use the ‘contingency table’?

FISCHERS EXACT TEST CONTINGENCY TABLE		
	<i>Positive</i>	<i>Negative</i>
<i>Chemo-sensitive</i>	x	x
<i>Chemo-resistance</i>	x	x

**Figure 24:** shows a  $2 \times 2$  contingency table for Fischer's exact test. The values (x) is entered for the number of patients that show a chemo-sensitive or chemo-resistant response against a positive or a negative IHC score for a particular staining localization.

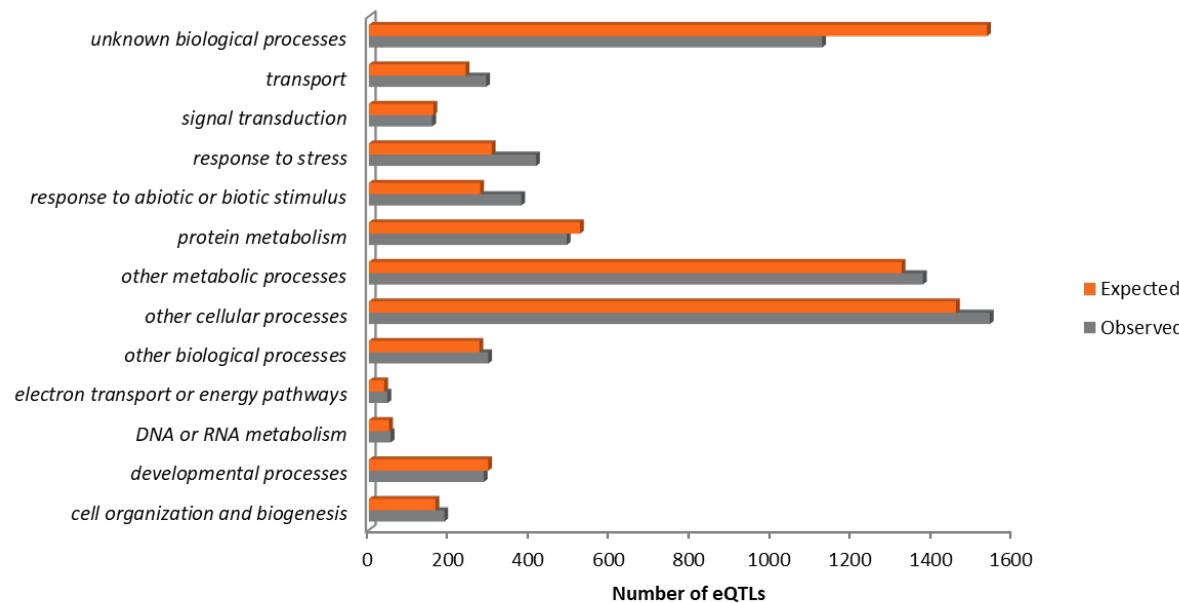
- 2 treatments x 2 accessions/strains
- 2 variables x 2 targets

## Ejemplo



	sRNA_1	sRNA_2
Planta Sin bicho	10	10
Planta Con bicho	10	30

## Gene Ontology Term enrichment Observed vs expected



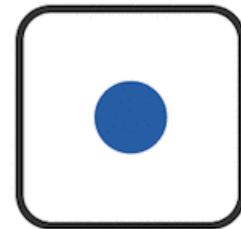
# Hypergeometric test



Binomial Test  
50/50

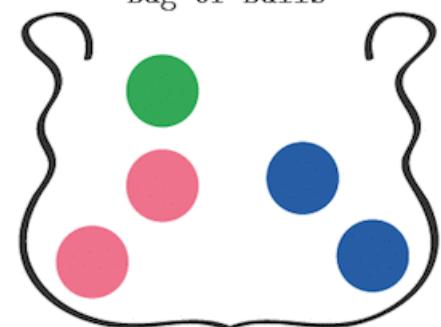
Stochastic Process

Random Variable



Possible States: ● ● ●

Bag of Balls



Hypergeometric test  
1/50

## Example

Universe: 18840 balls total  
red balls in the universe: 6680

Sample: 382 balls  
total red balls in the sample: 160

I would like to estimate if the percentage of red balls in my sample is significantly different from the percentage of reds in universe

`dhyper(x, m, n, k, log = FALSE)`

`x, q`

vector of quantiles representing the number of white balls drawn without replacement from an urn which contains both black and white balls.

`m`

the number of white balls in the urn.

`n`

the number of black balls in the urn.

`k`

the number of balls drawn from the urn.

`p`

probability, it must be between 0 and 1.

`dhyper(160, 6680, 12160, 382, log = FALSE)`

P = 0,013

## *Evidence for directional allelic effect*

